

Bartha-Vári J. H.¹, Nagy E. Z.¹, Gal C. A.¹, Bencze L. C.¹, Toşa M. I.¹, Irimie F. D.¹, Abaházi E.², Poppe L.^{2*}, Paizs C.^{1*}

¹ Department of Chemistry, Faculty of Chemistry and Chemical Engineering, Babeş-Bolyai University

² Department of Organic Chemistry and Technology, Budapest University of Technology and Economics

E-mail: paizs@chem.ubbcluj.ro, poppe@mail.bme.hu

Introduction

Numerous enantiopure heteroaryl alcohols are known for their biological activity, or represent precursors in the synthesis of a large number of pharmaceutical products.¹

An efficient method for the synthesis of enantiopure secondary alcohols is the lipase-catalyzed kinetic resolution of racemic alcohols.

Immobilization can modify and improve properties of enzymes, may enable their recovery and reuse, can provide a favorable environment for the enzymes, may increase their rigidity and may improve their enantioselectivity.²

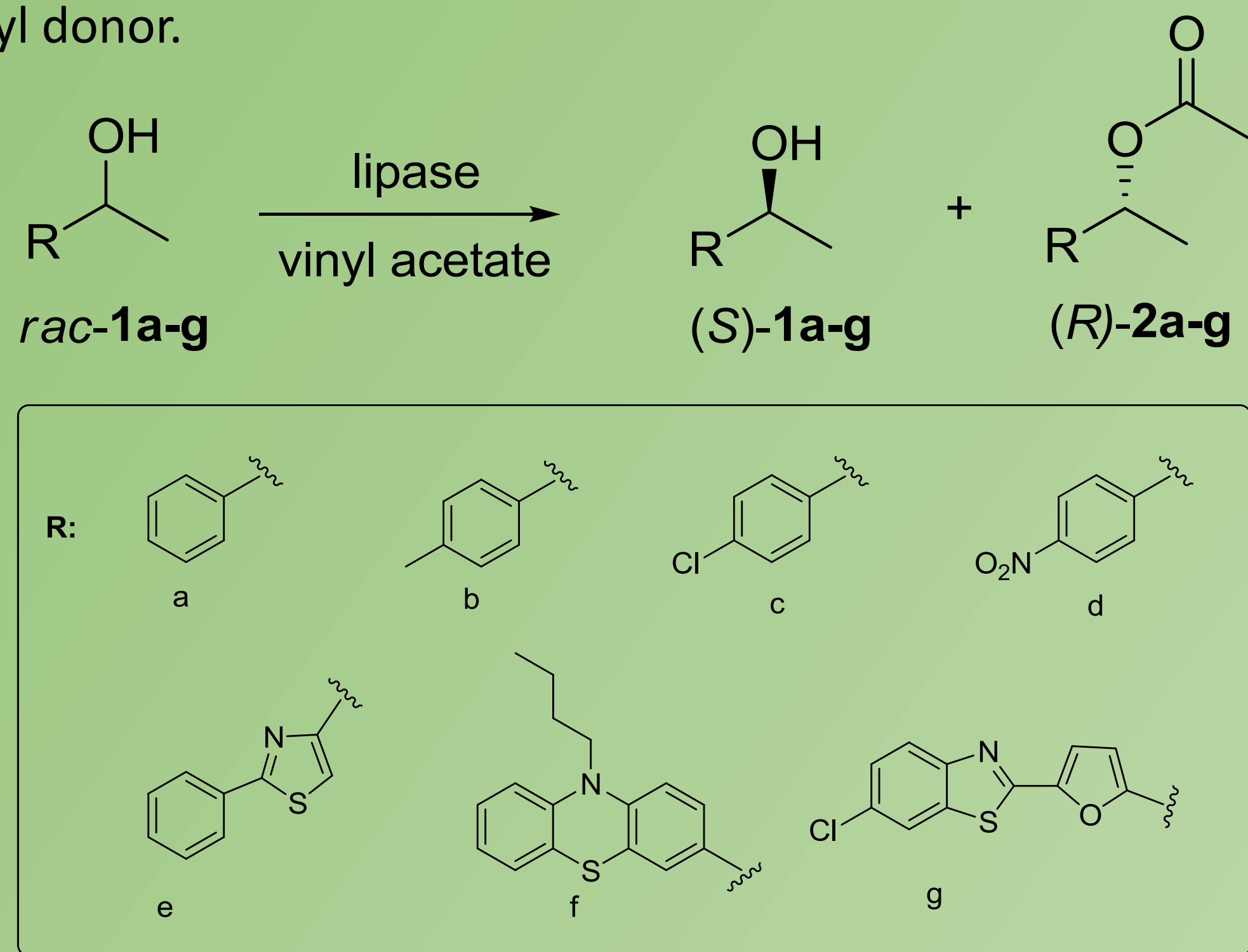
Carbon nanotubes are widely used for the immobilization of biomacromolecules, due to their mechanical, thermal, electrical properties and general biocompatibility since they can offer a support with large surface area, low diffusion limitation and easy recovery.³

Experimental

The commercially available SwCNT_{COOH} was activated with carbonyldiimidazol, followed by coupling with 1,3-diaminopropane. The SwCNT carrier was further treated with the glycerol diglycidyl ether crosslinker (Scheme 2). Immobilization of the lipase B from *Candida antarctica* (CaL-B) was carried out in the presence of Tween 80, a non-ionic surfactant. Furthermore, the optimal loading of the enzyme on the support material was also tested. The SwCNT_{COOH}CaL-B biocatalyst was used in the kinetic resolution of several secondary alcohols (Scheme 1).

Discussion

The obtained enzyme preparations were characterized by reproducibility and high immobilization yields (>99 % of the enzyme bound to SwCNT_{COOH}). The immobilized lipase showed high enantioselectivity and activity in the enzymatic kinetic resolution of the tested racemic secondary alcohols *rac-1a-g* (Table 1) in acetonitrile, using vinyl acetate as acyl donor.



Scheme 1. Enzymatic kinetic resolution of racemic alcohols *rac-1a-g*. Reactants and solvents: vinyl acetate, SwCNT_{COOH}CaL-B, acetonitrile.

Conclusions

Covalent immobilization of CaL-B on carboxylated single-walled carbon nanotubes after amidation and bisepoxide treatment resulted in a highly efficient and stable biocatalyst which proved to be useful in the kinetic resolution of various secondary alcohols.

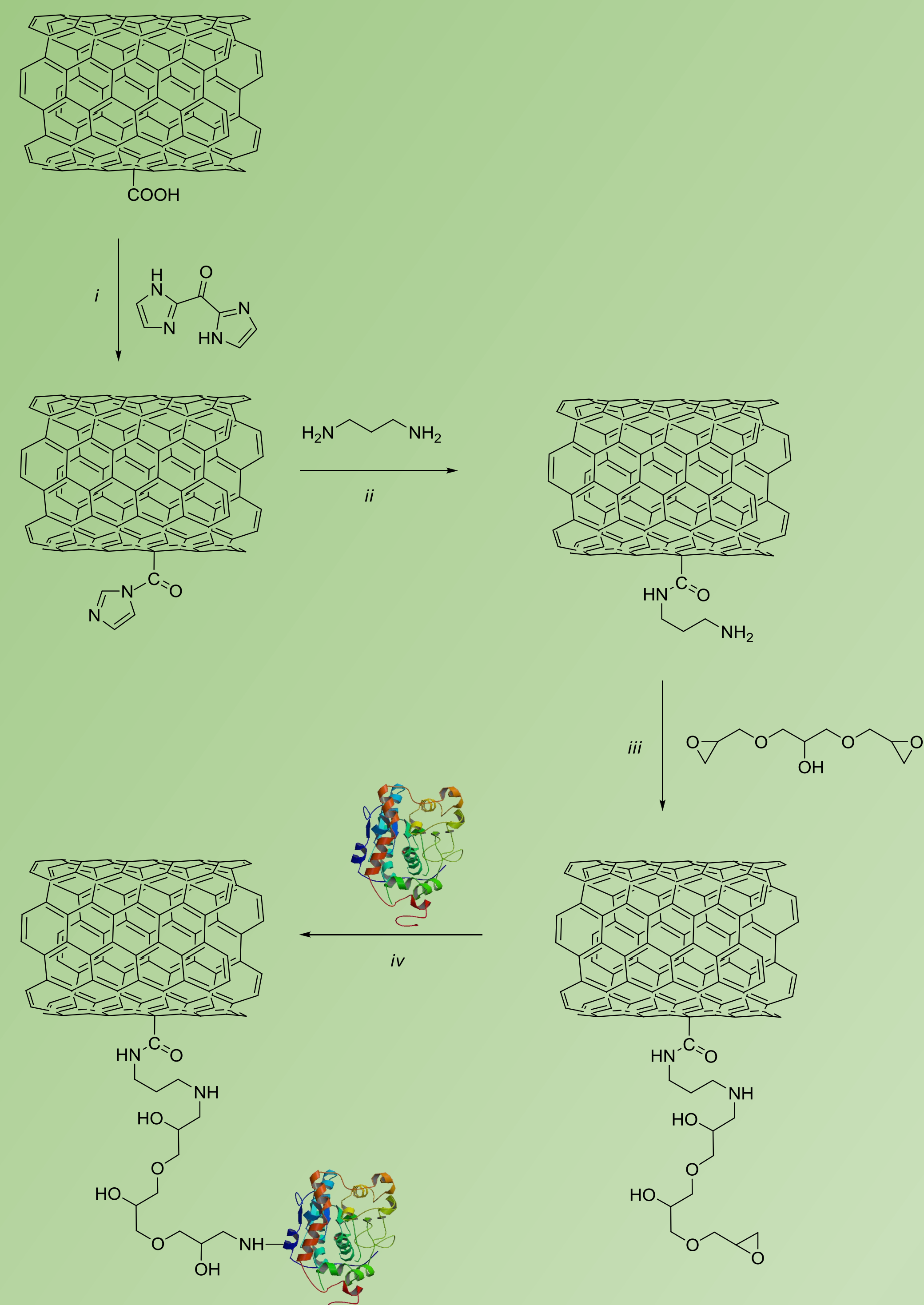
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Scheme 2. Immobilization of CaL-B on SwCNT_{COOH} i) CDI in CH₂Cl₂; ii) H₂N(CH₂)₃NH₂ in water; iii) glycerol diglycidyl ether in CH₂Cl₂; iv) CaL-B in PBS buffer (20 mM Na₂HPO₄, 150 mM NaCl, pH 7)

Table 1. Enantioselective acylation of *rac-1a-g* mediated by SwCNT_{COOH}CaL-B after 36 h (10 mg substrate, 1 mg neat enzyme, 5 µL vinyl acetate, 1 mL acetonitrile).

Substrate	<i>ee_s</i> (%)	<i>ee_p</i> (%)	<i>c</i> (%)	<i>E</i>
<i>rac-1a</i>	37	>99	27.2	>200
<i>rac-1b</i>	68	>99	40.6	>200
<i>rac-1c</i>	49	>99	33.1	>200
<i>rac-1d</i>	92	>99	48.0	»200
<i>rac-1e</i>	>99	>99	50	»200
<i>rac-1f</i>	>99	>99	50	»200
<i>rac-1g</i>	76	>99	43.1	»200

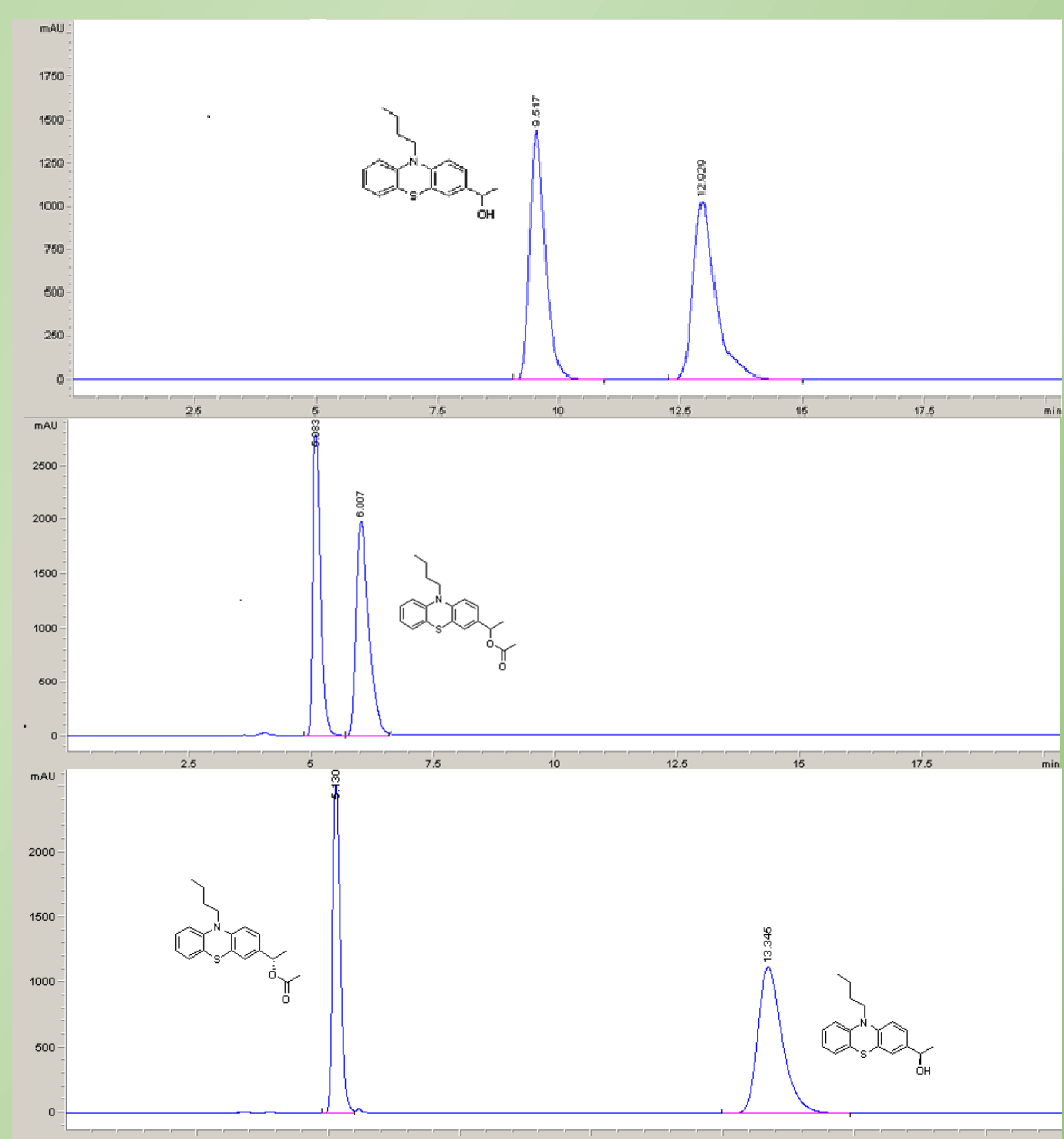


Figure 1. Chromatographic separation of the enantiomers of *rac-1f* and *rac-2f*; and enantiomeric composition of the products at 50% conversion in SwCNT_{COOH}CaL-B mediated enantiomer selective acylation of *rac-1f* with vinyl acetate in acetonitrile.